

Antibiotic therapy for pan-drug-resistant infections

Mahdi Asghari Ozma¹, Amin Abbasi², Mohammad Asgharzadeh³, Pasquale Pagliano⁴, Amedeo Guarino⁵, Şükran Köse⁶, Hossein Samadi Kafil⁷

¹Student Research Committee, Tabriz University of Medical Sciences, Tabriz, Iran;

²Department of Food Science and Technology, National Nutrition and Food Technology, Research Institute, Faculty of Nutrition Science and Food Technology, Shahid Beheshti University of Medical Sciences, Tehran, Iran;

³Biotechnology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran;

⁴Department of Medicine, University of Salerno, Salerno, Italy;

⁵Department of Public Health, University of Naples Federico II, Naples, Italy;

⁶Department of Infectious Diseases and Clinical Microbiology, 9 Eylül University, İzmir, Turkey;

⁷Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

Article received 1 November 2022, accepted 18 November 2022

SUMMARY

Antibiotic resistance occurs when microorganisms resist the drugs used against the infection caused by them and neutralize their effects over time using various mechanisms. These mechanisms include preventing drug absorption, changing drug targets, drug inactivating, and using efflux pumps, which ultimately cause drug resistance, which is named pan-drug-resistant (PDR) infection if it is resistant to all antimicrobial agents. This type of drug resistance causes many problems in society and faces the health system with difficulties; therefore their treatment is crucial and encourages doctors to develop new drugs to treat them. PDR Gram-negative bacteria, including *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Klebsiella pneumoniae*, and *Escherichia coli* are among the most significant resistant bacteria to many antimicrobial agents, and only a limited range of antibiotics, especially synergistically are effective on them. For the therapy of PDR *A. baumannii*, tigecycline in com-

ination with colestimethate, imipenem, amikacin, and ampicillin-sulbactam are the most effective treatments. The utilization of β -lactamase inhibitors such as ceftolozane-tazobactam, ceftazidime-avibactam, or imipenem-cilastatin-relebactam has the most efficacy against PDR *P. aeruginosa*. The PDR *K. pneumoniae* has been treated in the last decades with tigecycline and colistin, but currently, nitrofurantoin, fosfomycin, and pivmecillinam seem to be the most effective agent for the therapy of PDR *E. coli*. While these drugs impressively struggle with PDR pathogens, due to the daily increase in antibiotic resistance in microorganisms worldwide, there is still an urgent need for the expansion of novel medicines and methods of combating resistance.

Keywords: Antibiotic resistance, pan-drug-resistant, *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Klebsiella pneumoniae*, *Escherichia coli*, therapy.

INTRODUCTION

The emergence of resistance to various antimicrobial agents in bacteria has endangered human health worldwide, and the reason for this is the lack or absence of antimicrobial agents against

them [1, 2]. Antibiotic resistance exists in both Gram-positive and Gram-negative bacteria, and this problem has led to the emergence of new scientific terms in the field of antibiotic resistance [3, 4]. One of these definitions is multidrug-resistant organisms (MDR), which is the resistance of an organism to more than one antimicrobial agent, which makes the treatment ineffective against the infection or delays the treatment and endangers the patient's health [5]. Bacteria categorized as extensively drug-resistant (XDR) are not only

Corresponding author
Hossein Samadi Kafil
E-mail: Kafilh@tbzmed.ac.ir

resistant to multiple antibacterial substances, but also have the inauspicious potential of being resistant to almost all authorized antimicrobials, making them epidemiologically important [6]. Initially, the term XDR was used in medical sciences to broadly explain drug-resistant *Mycobacterium tuberculosis* (XDR MTB) as resistant to the first-line drugs rifampicin and isoniazid, to a fluoroquinolone and at least one of three mentioned second-line drugs, including amikacin, kanamycin, or capreomycin [7].

Among the terms of drug resistance, pan-drug-resistance (PDR) refers to resistance to all antimicrobial compounds [8]. PDR bacterial infections, on the side of the abuse of broad-spectrum antibiotics in clinical utilization, pose a considerable public health menace due to the resistance of pathogenic microorganisms to various antimicrobials [9]. As the problem progresses, these organisms show resistance to all presently utilized antibacterial substances or stay sensitive only to previously used and potentially more toxic agents such as polymyxins and tigecycline, leaving limited and non-optimal choices for the remedy of infections [10]. PDR Gram-negative bacteria, including *Pseudomonas aeruginosa*, *Acinetobacter baumannii*, *Klebsiella pneumoniae*, and *Escherichia coli* are among the most significant bacteria that show resistance to varied antibacterial substances, and only a limited range of antibiotics are effective on them [11]. These PDR bacteria are widely disseminated in the clinic because of their strong stability, colonization incidence rate, and resistance to antibacterial substances [12]. Since PDR bacteria adopt resistance mechanisms quickly, there is a debate about antibiotic choices concerning the efficacy and susceptibility of resistant strains in clinical utilization. Therefore, it is not easy to describe a standard therapeutic regimen against PDR infections in ill people [13]. The present narrative review will deal with the recent antibiotic advances in treating PDR infections and the most effective methods of treating them.

■ THE EMERGENCE OF ANTIBIOTIC RESISTANCE

Misuse and overuse of antimicrobial agents are the most significant causes of the expansion of drug-resistant microorganisms [14, 15]. Lack of clean water and sanitation and insufficient infec-

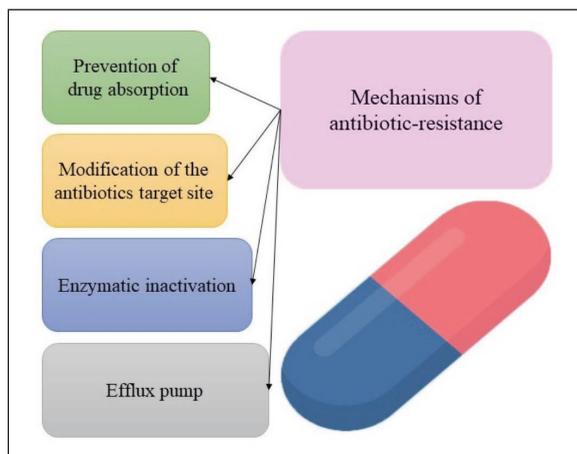


Figure 1 - The most significant mechanisms of antibiotic-resistance.

tion prohibition and control cause the dissemination of microorganisms, some of which can be resistant to antimicrobial remedies [16, 17]. Antimicrobial resistance mechanisms are classified into four main classes, *i.e.* preventing drug absorption, modifying the target of the drug, drug inactivating, and efflux pump. Intrinsic resistance is due to preventing drug absorption, drug inactivation, using an efflux pump, and, exerting the drug out of the cell. Acquired resistance is due to the modification of the target of drugs, enzymatic inactivation of drugs, and use of efflux pumps (Figure 1) [18]. There are differences in the kinds of mechanisms utilized by Gram-negative bacteria in comparison with Gram-positive ones, because of the diversity in bacterial structure [19]. All four principal mechanisms are utilized by Gram-negative bacteria, whereas Gram-positive bacteria just use preventing drug absorption [20, 21]. The existence of these differences causes variation in the drug resistance of different strains and each strain shows a different pattern of antibiotic resistance and causes variation in the effects of drugs on them [22].

■ TREATMENT OF PDR *A. BAUMANNII*

Acinetobacter baumannii, an aerobic, Gram-negative coccobacillus, is considered a crucial nosocomial pathogen because of various drug resistance mechanisms, and it can be a complicated microorganism for physicians to perform therapeutic

measures [23]. It causes multiple infectious disorders, including pneumonia, wound, urinary tract, hematological, and intra-abdominal infections [24]. The number of cases of *A. baumannii* with resistant patterns to carbapenems over a 2-year time is increased [25]. As expected, the emergence and dissemination of PDR *A. baumannii* isolate in hospital settings leads to high mortality rates and is difficult to eradicate [26]. The increasing rate of microorganisms has shown resistance to antibiotics such as carbapenems worldwide, which in past years had high efficacy in the therapy of patients [27]. Non-traditional substances such as polymyxin E and polymyxin B, are utilized for the therapy of patients with pan-drug-resistant *A. baumannii* despite their high toxicities [28-30]. Resistance to these antibacterial substances has also emerged among some strains during treatment with them [31].

Pharmacological measures with newer antibacterial drugs or antimicrobial synergistic combination utilization of drugs have become increasingly crucial in combating these infections. Tigecycline also was authorized for the therapy of complicated skin and gastrointestinal infections caused by susceptible microorganisms [32]. Treatment with tigecycline alone or in simultaneous consumption with other antibacterial substances including imipenem, colestimethate, amikacin, and ampicillin-sulbactam are impressive in treating PDR infection [33]. Another effective synergy was seen in the utilization of imipenem-colestimethate and tigecycline-imipenem combinations. These results demonstrate the importance of the use of synergistic consumption gaining the activity of particular antibacterial substance combinations against PDR *A. baumannii* [34].

■ TREATMENT OF PDR *P. AERUGINOSA*

P. aeruginosa is a ubiquitous bacterium that can survive under various environmental situations [35]. It not only causes disease in plants and animals but also in humans, causing multiple harsh disorders in immunocompromised patients with cancer and patients suffering from severe burns and cystic fibrosis (CF) [36]. This microorganism is one of the significant organisms responsible for drug-resistant nosocomial infections and is one of the crucial causes of biofilm formation, bacteremia, and pneumonia in hospitalized patients

[37, 38]. Multidrug-resistant *P. aeruginosa* was initially seen in patients with cystic fibrosis, and the spreading of them has since been announced among hospitalized patients [39]. *P. aeruginosa* isolates resistant to carbapenems or all antibacterial substances which are accessible for clinical utilization, are related to nosocomial infections and prevalence among patients hospitalized in intensive care units (ICUs) or burn units [40]. In addition to inherent resistance to multiple antimicrobial substances, *P. aeruginosa* acquires resistance quickly to conventional anti-pseudomonal agents such as carbapenems, ceftazidime, penicillins, fourth-generation cephalosporins, aztreonam, and ciprofloxacin following extended utilization of these antimicrobial agents in hospitalized patients, especially those in ICUs [41].

For the remedy of PDR *P. aeruginosa* infection, conventional, antipseudomonal β -lactam antibacterial drugs such as cefepime, ceftazidime, piperacillin-tazobactam, and carbapenems are generally prescribed, sometimes in simultaneous consumption with a second drug from another antibiotic class such as aminoglycoside, fluoroquinolone, polymyxin [42]. When the utilization of conventional β -lactams for *P. aeruginosa* infections may be ineffective due to the suspected PDR, monotherapy with novel β -lactamase inhibitors such as ceftolozane-tazobactam, ceftazidime-avibactam, or imipenem-cilastatin-relebactam over mix therapy with conventional substances is preferable, followed by therapeutic intensification or de-intensification based on *in vitro* susceptibility outcomes [43, 44]. If β -lactamase inhibitors are inaccessible, ceftiderocol is the preferable remedy for acute PDR *P. aeruginosa* infections over simultaneous consumption with conventional antibacterial substances [45-47].

■ TREATMENT OF PDR *K. PNEUMONIAE*

K. pneumoniae is a significant pathogen that often causes nosocomial infections in hospitalized patients. In the past decade, the clinical isolation rate of *K. pneumoniae* has been rising, and in most countries, it has become the second most prevalent clinically isolated Gram-negative bacilli after *E. coli* [48]. The emergence of carbapenem-resistant *K. pneumoniae* causes a complicated problem for public health due to the lack of effective therapeutic options for the therapy of such infections

[49, 50]. *K. pneumoniae* carbapenemase (KPC) is one of the most significant carbapenemases found in *K. pneumoniae*, whose acquisition has contributed to resistance to all β -lactams in this microorganism and the therapy of infections caused by them has faced many problems [51].

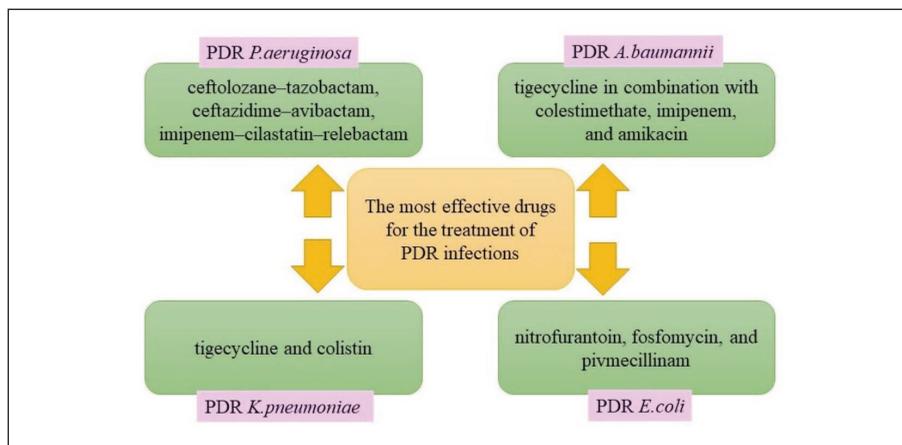
Carbapenem-resistant *K. pneumoniae* that has resistance to drugs such as tigecycline and colistin, often known as PDR bacteria or super bacteria, makes the therapy of the infections hard for clinicians [52]. PDR *K. pneumoniae* has been isolated in plenty of health centers of countries worldwide [53]. Most KPC-carrying *K. pneumoniae* strains are MDR and PDR microorganisms that have also resistance to β -lactams, aminoglycosides, fosfomicin, and quinolones [54]. Tigecycline and colistin are two effective antibacterial drugs with high efficacy for treating KPC-producing PDR *K. pneumoniae* infections [55]. Tigecycline was impressive against most KPC-producing *K. pneumoniae* at its initial clinical utilization, and colistin is presently identified as the last solution for the therapy of carbapenem-resistant *K. pneumoniae* strain infections [56, 57].

■ TREATMENT OF PDR *E. COLI*

E. coli, the type genus of the family Enterobacteriaceae, is the most representative facultative anaerobe microorganism in the human gastrointestinal system. Some strains have developed the capability to cause gastrointestinal, urinary, and central nervous system disorders [58]. Urinary tract infections (UTIs) caused by antibiotic-resistant *E.*

coli, especially PDR ones are a growing concern due to the unavailability of appropriate treatment options [59]. Knowledge of the common uropathogens, and local susceptibility patterns, is essential in determining suitable antibiotic therapy for UTIs [60]. The recommended first-line antibacterial substances utilized for acute uncomplicated cystitis in healthy nonpregnant females is a period of nitrofurantoin, fosfomicin tromethamine, or pivmecillinam [61]. High rates of resistance in microorganisms against drugs such as trimethoprim-sulfamethoxazole and ciprofloxacin prevent their utilization as effective remedies for UTIs in multiple countries, especially in patients who utilized them recently or in those who are in facing the infection of extended-spectrum β -lactamases (ESBLs)-producing *E. coli* [62]. Second-line choices include oral cephalosporins such as cephalexin or cefixime, fluoroquinolones, and β -lactams, such as amoxicillin-clavulanate. Running therapy choices for UTIs against *E. coli* with the ability to produce AmpC- β -lactamase include drugs such as fosfomicin, nitrofurantoin, pivmecillinam, fluoroquinolones, cefepime, piperacillin-tazobactam and carbapenems [63, 64]. Therapeutic choices for the UTIs caused by ESBLs-*E.coli* include nitrofurantoin, fosfomicin, pivmecillinam, amoxicillin-clavulanate, finafloxacin, and sitafloxacin, while pivmecillinam, fosfomicin, finafloxacin, and sitafloxacin are oral therapy choices for ESBLs- *K.pneumoniae* [65]. Because the drug resistance of the strains is increasing daily and microorganisms are developing resistance to the drugs used currently, the utilization of new antimicrobi-

Figure 2 - The most effective drugs for the treatment of PDR infections.



als is essential and it is wisely for the therapy of infections caused by drug-resistant organisms to avoid resistance and prevent the loss of effective therapies (Figure 2) [66].

■ CONCLUSIONS

Resistance among deadly Gram-negative pathogens has increased to epidemic proportions, especially within hospitals and acute care settings. Infections with drug-resistant microorganisms such as MDR, XDR, and PDR *E. coli*, *P. aeruginosa*, *K. pneumoniae*, and *A. baumannii* contribute to alarming high mortality rates in vulnerable populations and add healthcare costs through lengthening hospitalization, use of resources, lost productivity, and high acuity care needs. Addressing this problem requires both infection prevention and appropriate treatment. Knowledge of local patterns of resistance and individual risk factors for resistance will lead to better care for patients. Although there have been worldwide initiatives to expand new antibacterial substances against PDR Gram-negative pathogens, only limited advances have been made in the last few years. We are forced to rely on new combinations of old drugs largely, and our most impressive advances have been with new β -lactamase inhibitors such as avibactam, vaborbactam, and relebactam in simultaneous consumption with old cephalosporins and carbapenems. While these drugs combat PDR pathogens effectively, there is still a consecutive requirement to develop new medicines and methods of combating resistance.

Conflict of interest

None

Funding

This study was supported by Tabriz University of Medical Sciences with grant number 70277 and approved by the local ethic committee.

Acknowledgments

We thank all comments and helps by our colleagues from DARC center.

■ REFERENCES

[1] Ozma MA, Khodadadi E, Rezaee MA, et al. Bacterial proteomics and its application in pathogenesis studies. *Curr Pharm Biotechnol.* 2022; 23 (10), 1245-1256.

[2] Azargun R, Sadeghi MR, Soroush Barhaghi MH, et al. The prevalence of plasmid-mediated quinolone resistance and ESBL-production in Enterobacteriaceae isolated from urinary tract infections. *Infect Drug Resist.* 2018; 11, 1007-1014.

[3] Catalano A, Iacopetta D, Ceramella J, et al. Multi-drug resistance (MDR): A widespread phenomenon in pharmacological therapies. *Molecules.* 2022; 27 (3), 616.

[4] Bialvaei AZ, Kafil HS, Asgharzadeh M, et al. Current methods for the identification of carbapenemases. *J Chemother.* 2016; 28 (1), 1-19.

[5] Ozma MA, Lahouty M, Abbasi A, et al. Effective Bacterial Factors Involved in the Dissemination of Tuberculosis. 2022.

[6] Pillay S, Steingart KR, Davies GR, et al. Xpert MTB/XDR for detection of pulmonary tuberculosis and resistance to isoniazid, fluoroquinolones, ethionamide, and amikacin. *Cochrane Database of Systematic Reviews* 2022; (5).

[7] Ozma MA, Rashedi J, Poor BM, et al. Tuberculosis and diabetes mellitus in Northwest of Iran. *Infecti Disord Drug Targets.* 2020; 20 (5), 667-671.

[8] Ferry T, Kolenda C, Laurent F, et al. Personalized bacteriophage therapy to treat pandrug-resistant spinal *Pseudomonas aeruginosa* infection. *Nat Commun.* 2022; 13 (1), 1-6.

[9] Karakonstantis S, Kritsotakis EI, Gikas A. Pandrug-resistant Gram-negative bacteria: a systematic review of current epidemiology, prognosis and treatment options. *J Antimicrob Chemother.* 2020; 75 (2), 271-282.

[10] Karakonstantis S, Ioannou P, Kofteridis DD. In search for a synergistic combination against pandrug-resistant *A. baumannii*; methodological considerations. *Infection.* 2022; 50 (3), 569-581. doi: 10.1007/s15010-021-01748-w.

[11] Karakonstantis S, Ioannou P, Samonis G, et al. Systematic review of antimicrobial combination options for pandrug-resistant *Acinetobacter baumannii*. *Antibiotics.* 2021; 10 (11), 1344.

[12] Addis T, Araya S, Desta K. Occurrence of multiple, extensive and pan drug-resistant *pseudomonas aeruginosa* and carbapenemase production from presumptive isolates stored in a Biobank at Ethiopian Public Health Institute. *Infect Drug Res.* 2021; 14, 3609.

[13] Bassetti M, Righi E, Vena A, et al. Risk stratification and treatment of ICU-acquired pneumonia caused by multidrug-resistant/extensively drug-resistant/pandrug-resistant bacteria. *Curr Opin Crit Care.* 2018; 24 (5), 385-393.

[14] Karakonstantis S, Kalemaki D. Antimicrobial overuse and misuse in the community in Greece and link to antimicrobial resistance using methicillin-resistant *S. aureus* as an example. *J Infect Public Health* 2019; 12 (4), 460-464.

[15] Ozma MA, Khodadadi E, Rezaee MA, et al. Induction of proteome changes involved in biofilm formation of *Enterococcus faecalis* in response to gentamicin. *Microb Pathog.* 2021; 157, b105003.

- [16] Dadgostar P. Antimicrobial resistance: implications and costs. *Infecti Drug Res* 2019; 12, 3903.
- [17] Abushaheen MA, Fatani AJ, Alosaimi M, et al. Antimicrobial resistance, mechanisms and its clinical significance. *Dis Mon.* 2020; 66 (6), 100971.
- [18] Sierra JM, Viñas M. Future prospects for Antimicrobial peptide development: Peptidomimetics and antimicrobial combinations. *Expert Opin Drug Discov.* 2021; 16 (6), 601-604.
- [19] Shi X, Xia Y, Wei W, et al. Accelerated spread of antibiotic resistance genes (ARGs) induced by non-antibiotic conditions: roles and mechanisms. *Water Res.* 2022; 224, 119060. doi: 10.1016/j.watres.2022.119060.
- [20] Subramaniam G, Girish M. Antibiotic resistance - a cause for reemergence of infections. *The Indian J Ped.* 2020; 87 (11), 937-944.
- [21] Kafil HS, Mobarez AM. Assessment of biofilm formation by enterococci isolates from urinary tract infections with different virulence profiles. *J King Saud Univ Sci.* 2015; 27 (4), 312-317.
- [22] Mobarki N, Almerabi B, Hattan A. Antibiotic resistance crisis. *Int J Med Dev Ctries* 2019; 40 (4), 561-564.
- [23] Kanaan MHG, Khashan HT. Molecular typing, virulence traits and risk factors of pandrug-resistant *Acinetobacter baumannii* spread in intensive care unit centers of Baghdad city, Iraq. *Revi Med Microbiol.* 2022; 33 (1), 51-55.
- [24] Cherubini S, Perilli M, Segatore B, et al. Whole-genome sequencing of ST2 *A. baumannii* causing bloodstream infections in COVID-19 patients. *Antibiotics*, 2022; 11 (7), 955.
- [25] Whiteway C, Breine A, Philippe C, et al. *Acinetobacter baumannii*. *Trends Microbiol.* 2022; 30 (2), 199-200. doi: 10.1016/j.tim.2021.11.008.
- [26] Antunes L, Visca P, Towner KJ. *Acinetobacter baumannii*: evolution of a global pathogen. *Pathog Dis.* 2014; 71 (3), 292-301. doi: 10.1111/2049-632X.12125.
- [27] Göttig S, Gruber TM, Higgins PG, et al. Detection of pan drug-resistant *Acinetobacter baumannii* in Germany. *J Antimicrob Chemother.* 2014; 69 (9), 2578-2579.
- [28] Valcek A, Nesporova K, Whiteway C, et al. Genomic analysis of a strain collection containing multidrug-, extensively drug-, pandrug-, and carbapenem-resistant modern clinical isolates of *Acinetobacter baumannii*. *Antimicrobial Agents Chemother.* 2022; 66 (9), e00892-00822.
- [29] Vahhabi A, Hasani A, Rezaee MA, et al. A plethora of carbapenem resistance in *Acinetobacter baumannii*: No end to a long insidious genetic journey. *J Chemother.* 2021; 33 (3), 137-155.
- [30] Kannian P, Mahanathi P, Ashwini V, et al. Carbapenem-Resistant Gram negative bacilli are predominantly multidrug or pan-drug resistant. *Microbial Drug Resistance* 2021; 27(8), 1057-1062.
- [31] Lim CLL, Chua AQ, Teo JQM, et al. Importance of control groups when delineating antibiotic use as a risk factor for carbapenem resistance, extreme-drug resistance, and pan-drug resistance in *Acinetobacter baumannii* and *Pseudomonas aeruginosa*: A systematic review and meta-analysis. *Intern J Infect Dis.* 2018; 76, 48-57.
- [32] Fragkou PC, Poulakou G, Blizou A, et al. The role of minocycline in the treatment of nosocomial infections caused by multidrug, extensively drug and pandrug resistant *Acinetobacter baumannii*: a systematic review of clinical evidence. *Microorganisms.* 2019; 7 (6), 159.
- [33] Karakonstantis S. A systematic review of implications, mechanisms, and stability of in vivo emergent resistance to colistin and tigecycline in *Acinetobacter baumannii*. *J Chemother.* 2020; 33 (1), 1-11.
- [34] Deng Z-W, Wang J, Qiu C-F, et al. A case report of intraventricular and intrathecal tigecycline infusions for an extensively drug-resistant intracranial *Acinetobacter baumannii* infection. *Medicine* 2019; 98 (15).
- [35] Ciofu O, Tolker-Nielsen T. Tolerance and resistance of *Pseudomonas aeruginosa* biofilms to antimicrobial agents - how *P. aeruginosa* can escape antibiotics. *Front Microbiol.* 2019; 10, 913.
- [36] Rasamiravaka T, El Jaziri M. Quorum-sensing mechanisms and bacterial response to antibiotics in *P. aeruginosa*. *Curr Microbiol.* 2016; 73 (5), 747-753.
- [37] Raman G, Avendano EE, Chan J, et al. Risk factors for hospitalized patients with resistant or multidrug-resistant *Pseudomonas aeruginosa* infections: a systematic review and meta-analysis. *Antimicrob Resist Infect Control.* 2018; 7 (1), 1-14. doi: 10.1186/s13756-018-0370-9.
- [38] Ozma MA, Khodadadi E, Pakdel F, et al. Baicalin, a natural antimicrobial and anti-biofilm agent. *J Herbal Med.* 2021; 27: 100432.
- [39] Vaez H, Salehi-Abargouei A, Ghalehnoo ZR, et al. Multidrug resistant *Pseudomonas aeruginosa* in Iran: A systematic review and metaanalysis. *J Glob Infect Dis.* 2018; 10 (4), 212.
- [40] Bassetti M, Castaldo N, Cattelan A, et al. Ceftolozane/tazobactam for the treatment of serious *Pseudomonas aeruginosa* infections: a multicentre nationwide clinical experience. *Int J Infect Dis.* 2019; 53 (4), 408-415.
- [41] Gorityala BK, Guchhait G, Goswami S, et al. Hybrid antibiotic overcomes resistance in *P. aeruginosa* by enhancing outer membrane penetration and reducing efflux. *J Med Chem.* 2016; 59 (18), 8441-8455.
- [42] Samad T, Co JY, Witten J, et al. Mucus and mucin environments reduce the efficacy of polymyxin and fluoroquinolone antibiotics against *Pseudomonas aeruginosa*. *ACS Biomaterials Science & Engineering.* 2019; 5 (3), 1189-1194.
- [43] von Silva-Tarouca MS, Wolf G, Mueller RS. Determination of minimum inhibitory concentrations for silver sulfadiazine and other topical antimicrobial agents against strains of *Pseudomonas aeruginosa* isolated from canine otitis externa. *Vet Dermatol.* 2019; 30 (2), 145-e142.
- [44] Zheng X, Cao Q, Cao Q, et al. Discovery of syner-

- gistic activity of fluoroquinolones in combination with antimicrobial peptides against clinical polymyxin-resistant *Pseudomonas aeruginosa* DK2. *Chinese Chemical Letters*. 2020; 31 (2), 413-417.
- [45] Tamma PD, Aitken SL, Bonomo RA, et al. Infectious Diseases Society of America guidance on the treatment of extended-spectrum β -lactamase producing Enterobacterales (ESBL-E), carbapenem-resistant Enterobacterales (CRE), and *Pseudomonas aeruginosa* with difficult-to-treat resistance (DTR-P. *aeruginosa*). *Clin Infect Dis*. 2021; 72 (7), e169-e183.
- [46] Saki M, Farajzadeh Sheikh A, Seyed-Mohammadi S, et al. Occurrence of plasmid-mediated quinolone resistance genes in *Pseudomonas aeruginosa* strains isolated from clinical specimens in southwest Iran: a multi-central study. *Sci Reports*. 2022; 12 (1), 1-8.
- [47] Ulloa ER, Sakoulas G. Azithromycin: An underappreciated quinolone-sparing oral treatment for *Pseudomonas aeruginosa* infections. *Antibiotics*. 2022; 11 (4), 515.
- [48] Russo TA, Olson R, Fang C-T, et al. Identification of biomarkers for differentiation of hypervirulent *Klebsiella pneumoniae* from classical *K. pneumoniae*. *J Clin Microbiol*. 2018; 56 (9), e00776-00718.
- [49] Victor LY, Hansen DS, Ko WC, et al. Virulence characteristics of *Klebsiella* and clinical manifestations of *K. pneumoniae* bloodstream infections. *Emerg Infect Dis*. 2007; 13 (7), 986.
- [50] Ozma MA, Abbasi A, Ahangarzadeh Rezaee M, et al. A critical review on the nutritional and medicinal profiles of garlic's (*Allium sativum* L.) Bioactive Compounds. *Food Reviews International*. 2022; 1-38.
- [51] Campos AC, Albiero J, Ecker AB, et al. Outbreak of *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*: a systematic review. *Am J Infect Contr*. 2016; 44 (11), 1374-1380.
- [52] Zhang Y, Guo L-Y, Song W-Q, et al. Risk factors for carbapenem-resistant *K. pneumoniae* bloodstream infection and predictors of mortality in Chinese paediatric patients. *BMC Infect Dis*. 2018; 18 (1), 1-10.
- [53] Gasink LB, Edelstein PH, Lautenbach E, et al. Risk factors and clinical impact of *Klebsiella pneumoniae* carbapenemase-producing *K. pneumoniae*. *Infect Contr Hosp Epidemiol*. 2009; 30 (12), 1180-1185.
- [54] El-Badawy MF, El-Far SW, Althobaiti SS, et al. The first Egyptian report showing the co-existence of blaNDM-25, blaOXA-23, blaOXA-181, and blaGES-1 among Carbapenem-Resistant *K. pneumoniae* clinical isolates genotyped by BOX-PCR. *Infect Drug Res*. 2020; 13, 1237.
- [55] Ruan Z, Wu J, Chen H, et al. Hybrid genome assembly and annotation of a pandrug-resistant *Klebsiella pneumoniae* strain using nanopore and illumina sequencing. *Infect Drug Res*. 2020; 13, 199.
- [56] Alghoribi MF, Alqurashi M, Okdah L, et al. Successful treatment of infective endocarditis due to pandrug-resistant *Klebsiella pneumoniae* with ceftazidime-avibactam and aztreonam. *Sci Rep*. 2021; 11 (1), 1-9.
- [57] Xu J, Zhao Z, Ge Y, et al. Rapid emergence of a pandrug-resistant *Klebsiella pneumoniae* ST11 isolate in an inpatient in a teaching hospital in China after treatment with multiple broad-spectrum antibiotics. *Infect Drug Res*. 2020; 13, 799.
- [58] Azam M, Jan AT, Kumar A, et al. Study of pandrug and heavy metal resistance among *E. coli* from anthropogenically influenced Delhi stretch of river Yamuna. *Braz J Microbiol*. 2018; 49, 471-480.
- [59] Nkansa-Gyamfi NA, Kazibwe J, Traore DA, et al. Prevalence of multidrug-, extensive drug-, and pandrug-resistant commensal *Escherichia coli* isolated from healthy humans in community settings in low- and middle-income countries: a systematic review and meta-analysis. *Glob Health Action*. 2019; 12 (Suppl. 1), 1815272.
- [60] Kim J, Hwang BK, Choi H, et al. Characterization of mcr-1-harboring plasmids from pan drug-resistant *Escherichia coli* strains isolated from retail raw chicken in South Korea. *Microorganisms*. 2019; 7n (9), n344.
- [61] Benklaouz MB, Aggad H, Benameur Q. Resistance to multiple first-line antibiotics among *Escherichia coli* from poultry in Western Algeria. *Vet World*. 2020; 13n (2), 290.
- [62] Feuerstein A, Scuda N, Klose C, et al. Antimicrobial resistance, serologic and molecular characterization of *E. coli* isolated from calves with severe or fatal enteritis in Bavaria, Germany. *Antibiotics*. 2021; 11 (1), 23.
- [63] Poirel L, Kieffer N, Liassine N, et al. Plasmid-mediated carbapenem and colistin resistance in a clinical isolate of *Escherichia coli*. *The Lancet Infect Dis*. 2016; 16 (3), 281.
- [64] Yao X, Doi Y, Zeng L, et al. Carbapenem-resistant and colistin-resistant *Escherichia coli* co-producing NDM-9 and MCR-1. *The Lancet Infect Dis*. 2016; 16 (3), 288-289.
- [65] Page MG, Bush K. Discovery and development of new antibacterial agents targeting Gram-negative bacteria in the era of pandrug resistance: is the future promising? *Curr Opin Pharmacol*. 2014; 18, 91-97.
- [66] Falagas ME, Mavroudis AD, Vardakas KZ. The antibiotic pipeline for multi-drug resistant gram negative bacteria: what can we expect? *Expert Rev Anti Infect Ther*. 2016; 14 (8), 747-763.